

A 3D ribbon diagram of a protein structure, colored in green, blue, and orange. The protein is shown in a ribbon representation, with a ligand molecule (represented by red, white, and blue spheres) bound to it. The background is black.

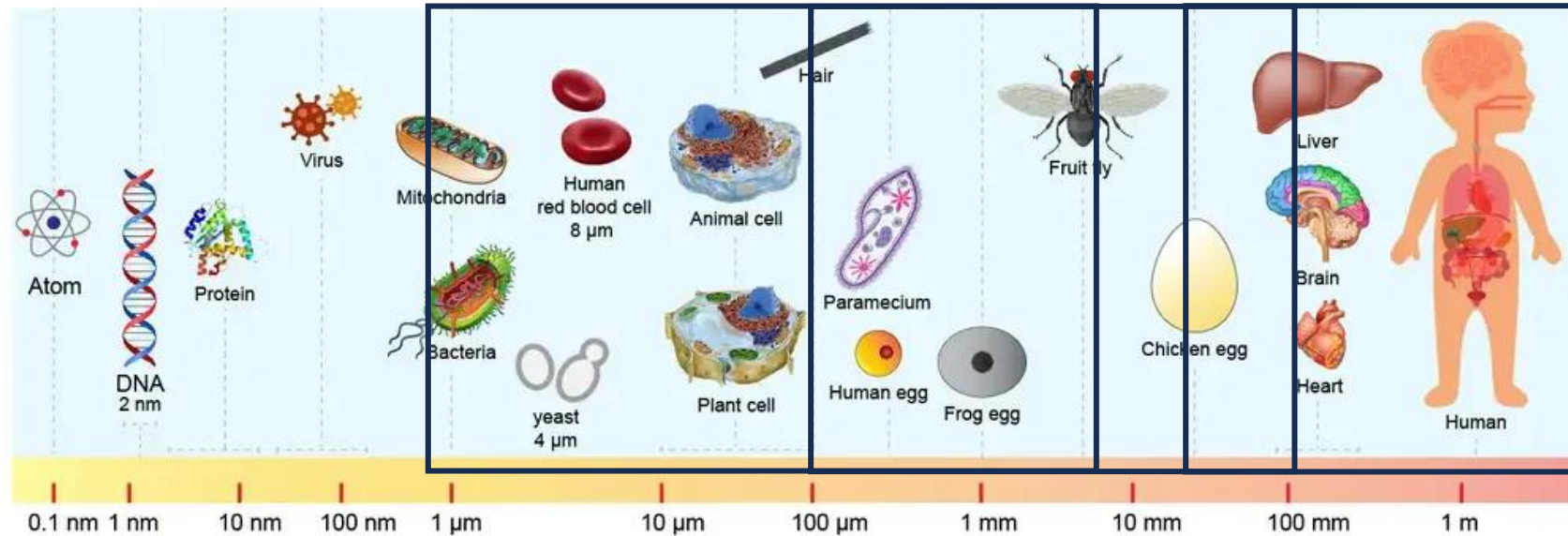
# Protein-based optical sensors for bioimaging applications

Takuya TERAI

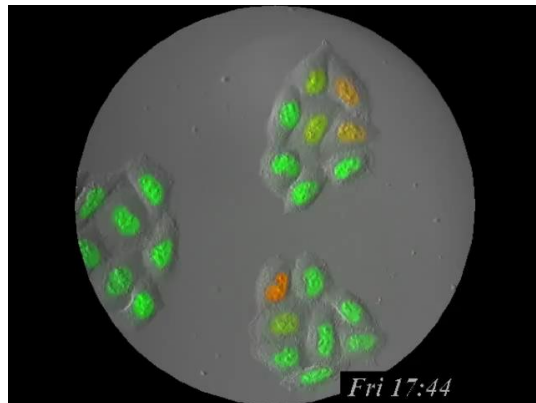
Department of Chemistry, School of Science,  
The University of Tokyo

# Bio(medical) imaging

## What we want to see in biology



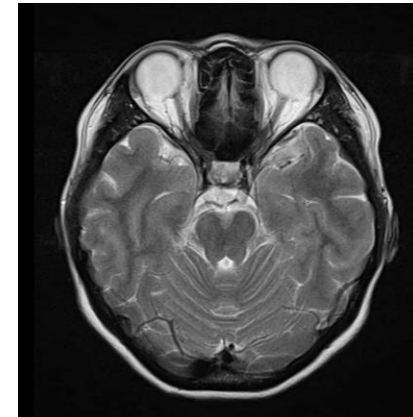
Fluorescence (cell)



Bioluminescence (mouse)



MRI (human brain)



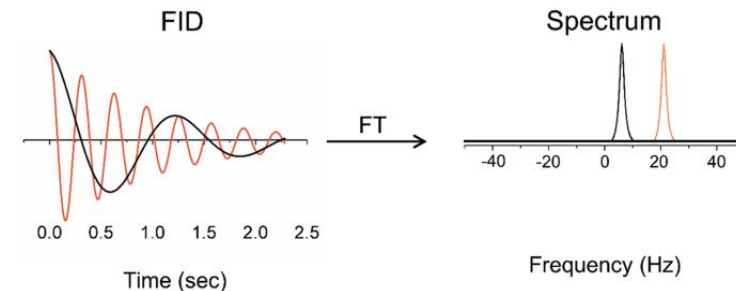
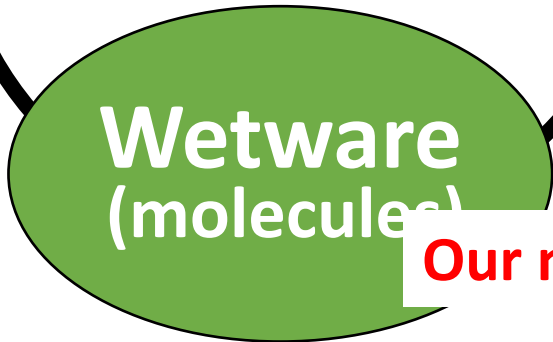
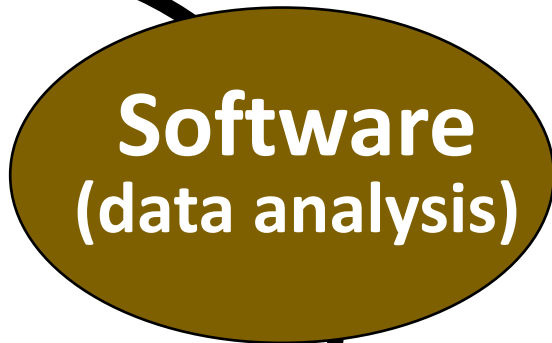
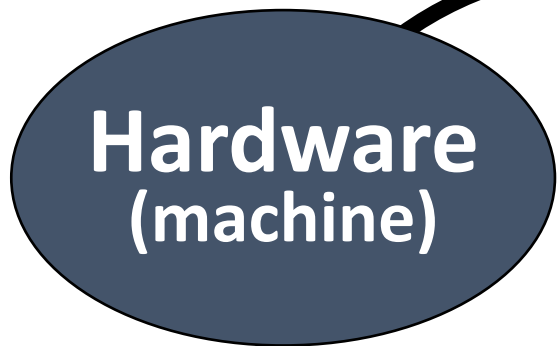
# Three components for bioimaging



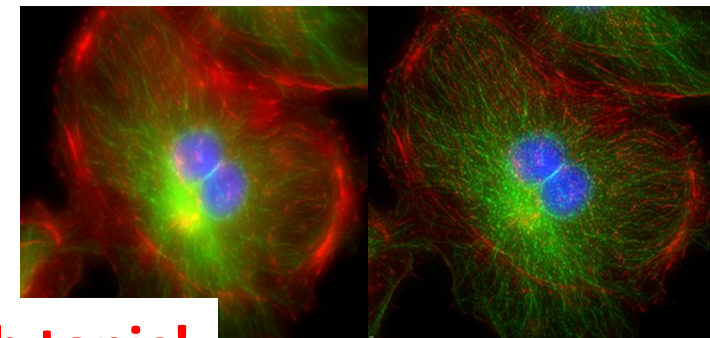
Fluorescence microscope



MRI@ETHZ

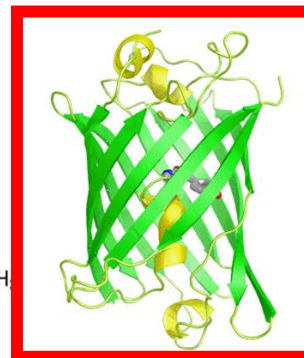
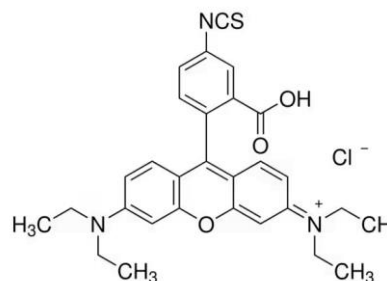
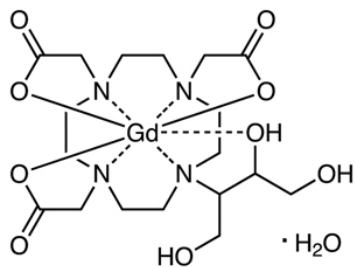


Fourier transformation of MR signal



Our main research topic! on of fluorescence images

Gd<sup>3+</sup> contrast agent  
(for MRI)

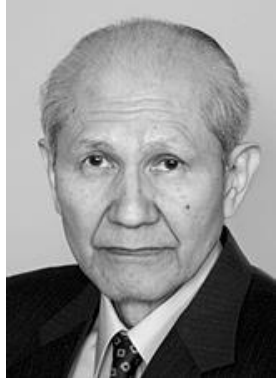


Fluorescent molecules/proteins  
(for fluorescence imaging)



# Discovery and application of GFP

In 2008, the Nobel prize in Chemistry was awarded to the three scientists for “the discovery and development of the green fluorescent protein, GFP”



Osamu Shimomura

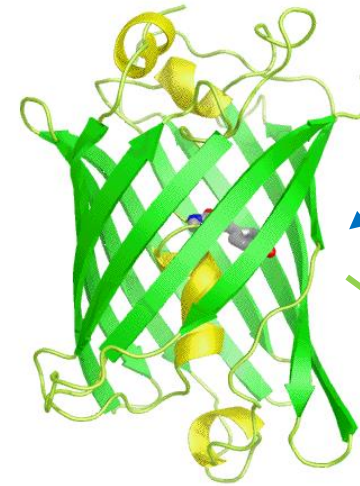


Martin Chalfie



Roger Y. Tsien

© The Nobel Foundation. Photo: U. Montan



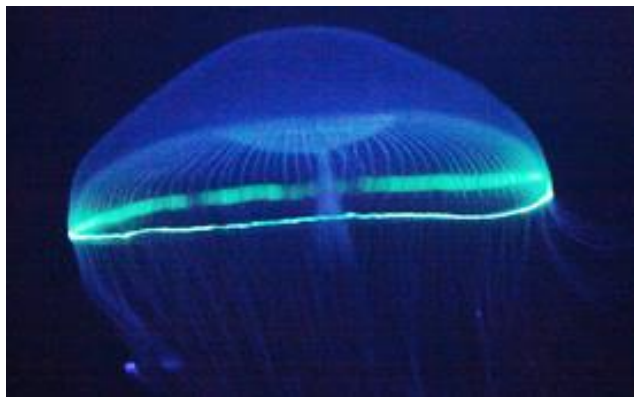
**Excitation light**

Ex. 395 nm (wt GFP)

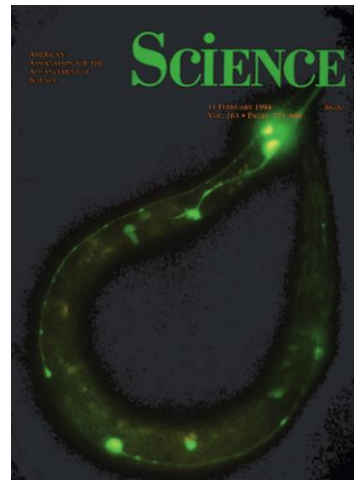
**Fluorescent light (emission)**

Em. 509 nm

GFP (PDB 1EMA)

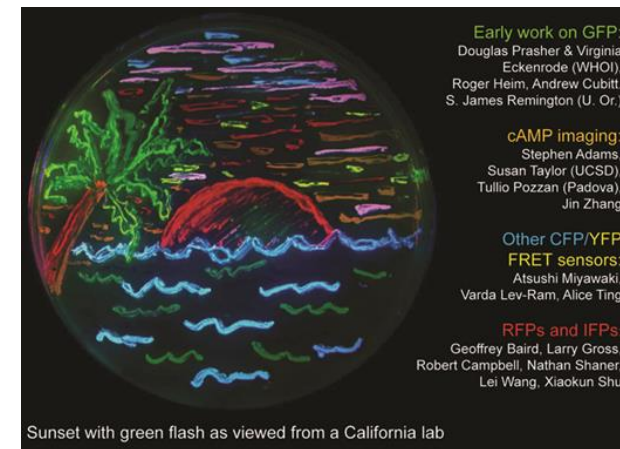


Fluorescent jerry fish  
(*Aequorea victoria*)



*C. elegans*  
expressing  
GFP

*Science*, 263, 802 (1994)



**Early work on GFP:**  
Douglas Prasher & Virginia Eckenrode (WHOI), Roger Heim, Andrew Cubitt, S. James Remington (U. Or.)

**cAMP imaging:**  
Stephen Adams, Susan Taylor (UCSD), Tullio Pozzan (Padova), Jin Zhang

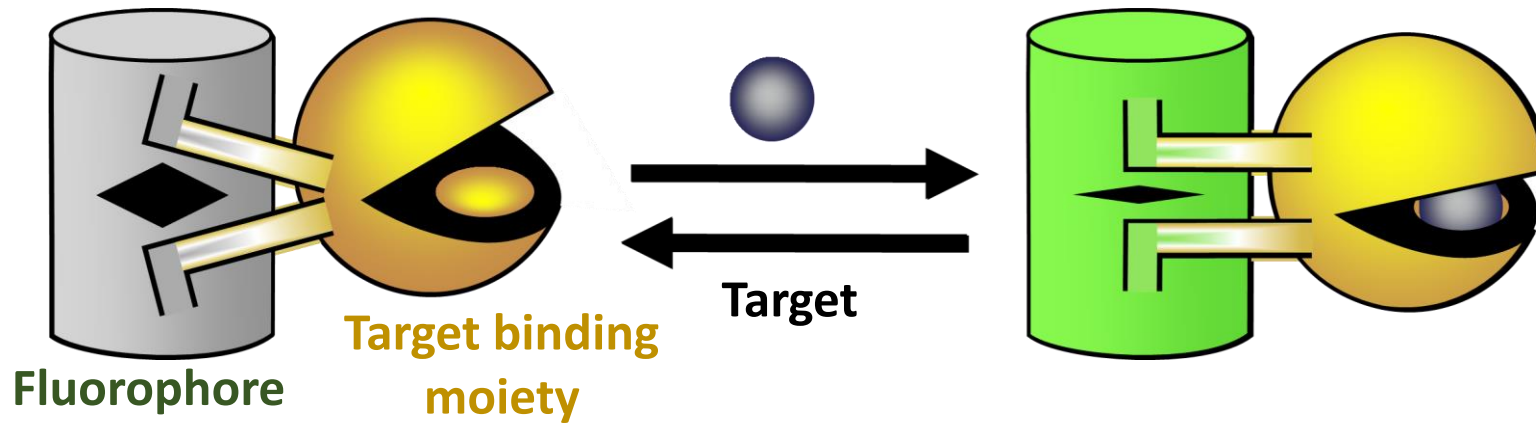
**Other CFP/YFP**  
**FRET sensors:**  
Atsushi Miyawaki, Varda Lev-Ram, Alice Ting

**RFPs and IFPs:**  
Geoffrey Baird, Larry Gross, Robert Campbell, Nathan Shaner, Lei Wang, Xiaokun Shu

A palette of  
fluorescent  
proteins

# Fluorescent sensors for bioimaging

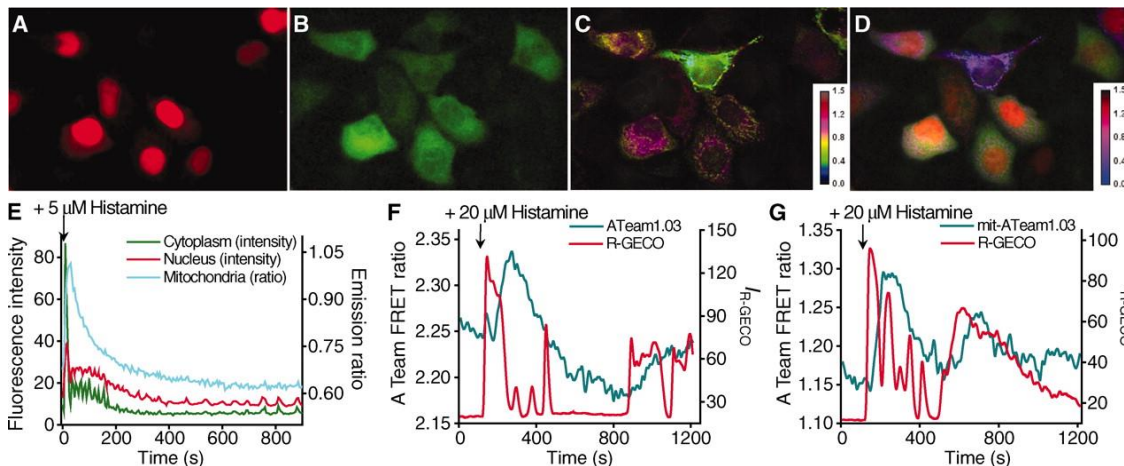
## Fluorescent probe (biosensor, indicator)



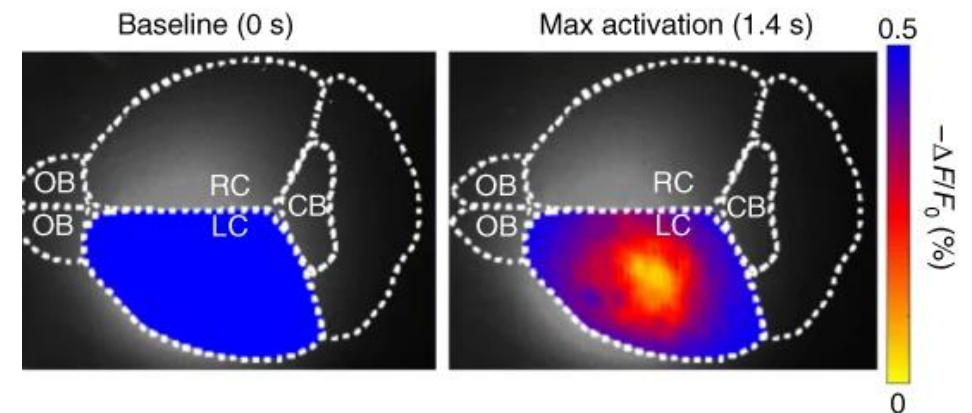
A molecule (synthetic, protein, or others) that reacts with the analyte and changes its fluorescence properties (intensity, wavelength).

## Ca<sup>2+</sup> imaging in cells/organisms

### B/G/R-GECO1

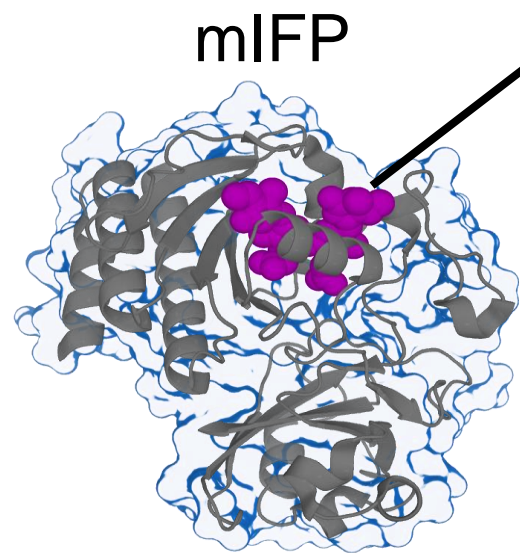
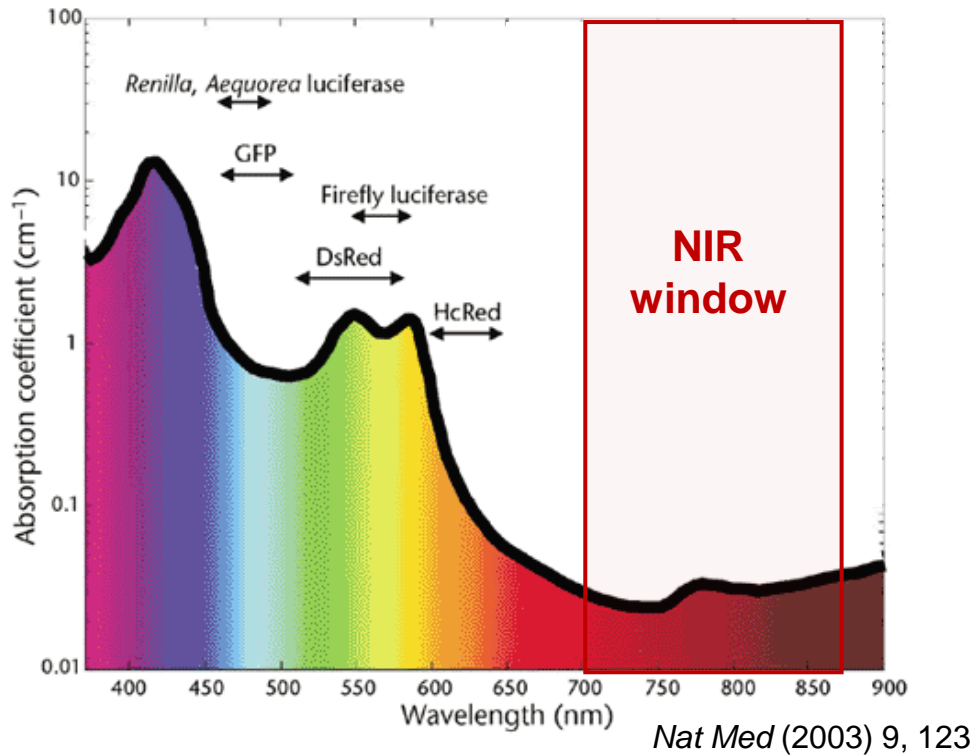


### NIR-GECO1

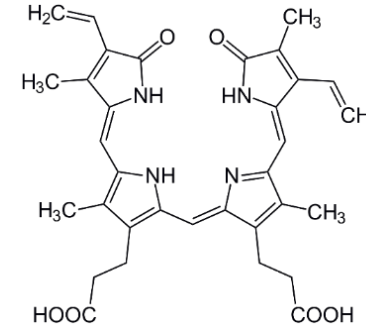


*Science*, 2011, 333, 1888; *Nat. Methods*, 2019, 16, 171.

# Near infrared (NIR) Ca<sup>2+</sup> sensors



## Biliverdin (BV)

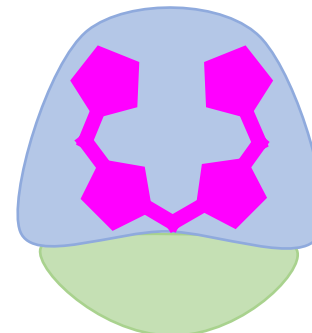


*Nat Methods*, 2015, 12, 763

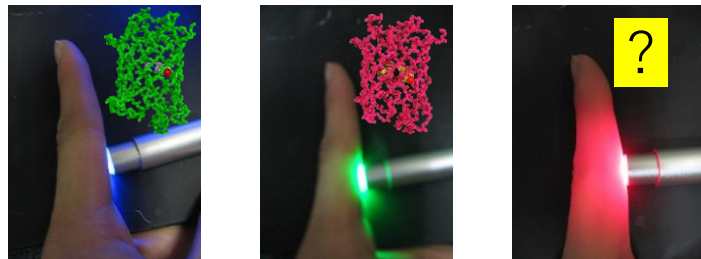
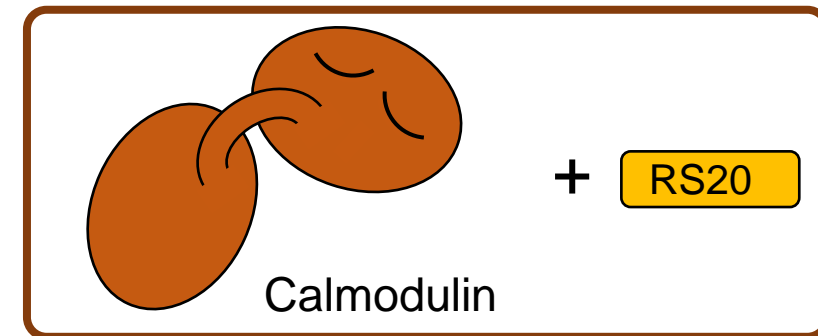
- Ubiquitous in mammalian tissues
- An intermediate in *heme* catabolism

$\epsilon = 82,000 \text{ M}^{-1}\text{cm}^{-1}$ ;  $\phi = 8\%$   
Ex 683 nm; Em 704 nm

||



## Target binding moiety



Short WL  
(blue)

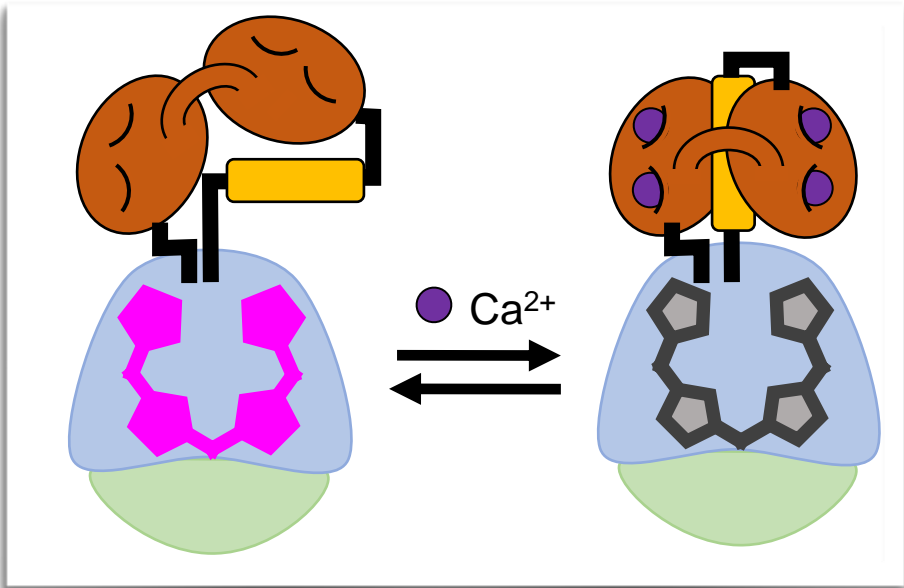


Long WL  
(red)

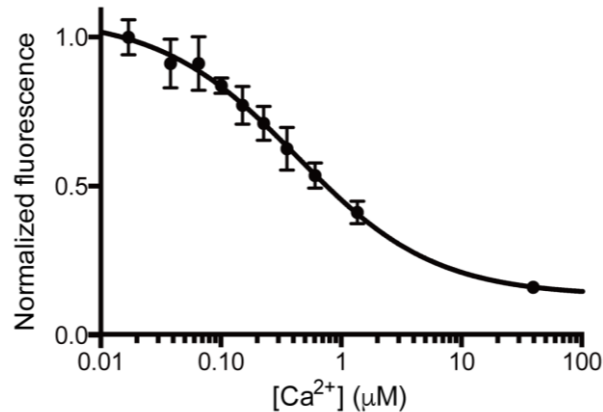
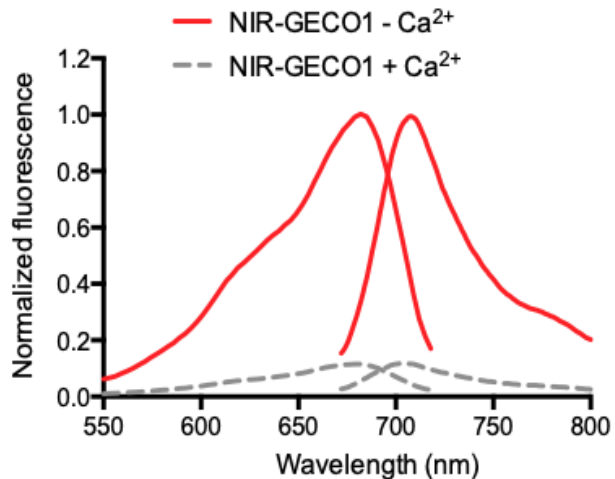
Fluorophore



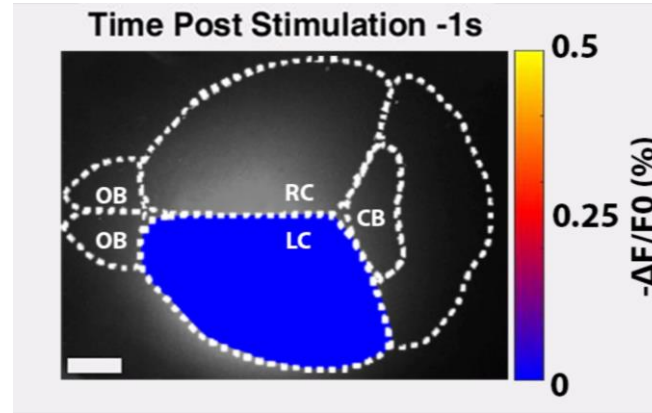
# Development of NIR-GECO1/2



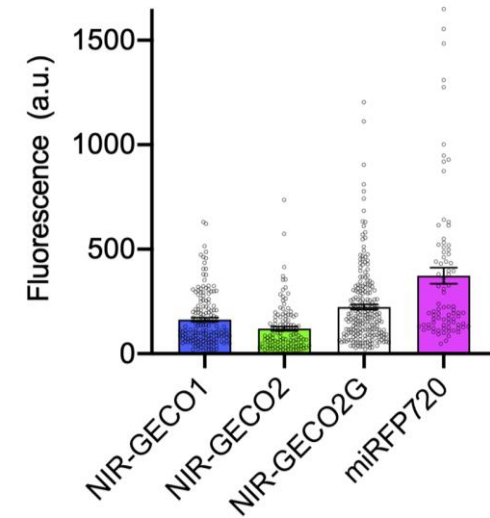
NIR-GECO2  $\lambda_{ex}$ : 678 nm,  $\lambda_{em}$ : 704 nm



## NIR-GECO1 in mouse brain



## Brightness of NIR-GECOs

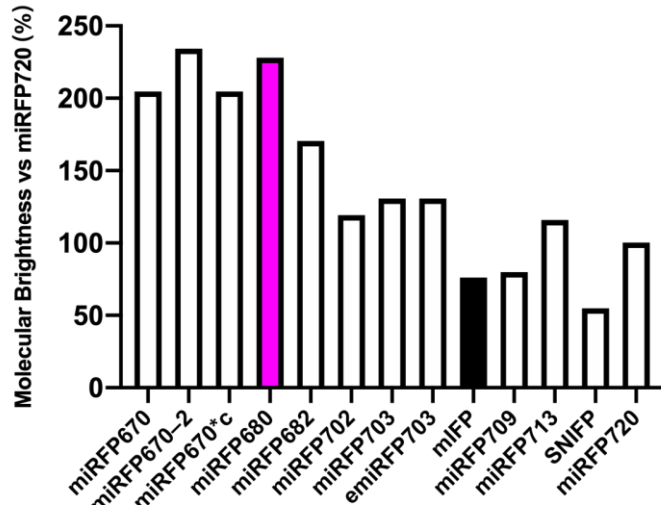


The sensor was not bright because

1. mIFP is dim itself.
2. Affinity to BV needs improvement.

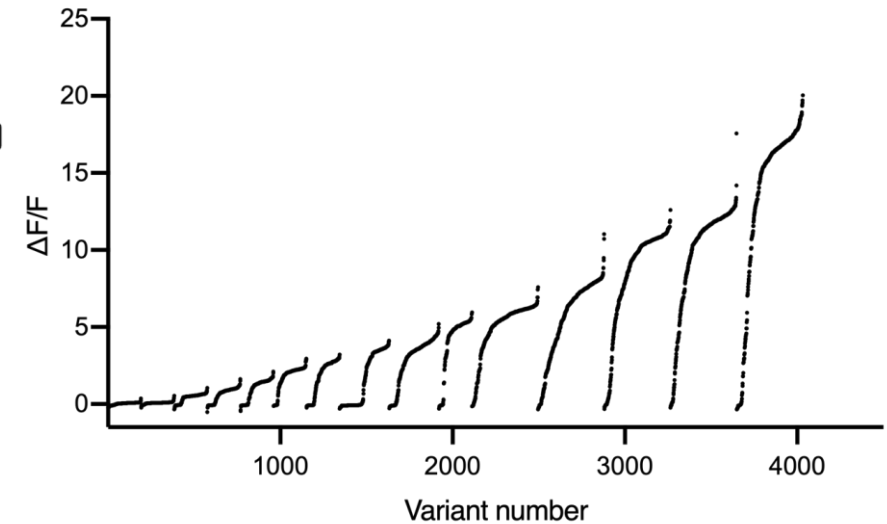
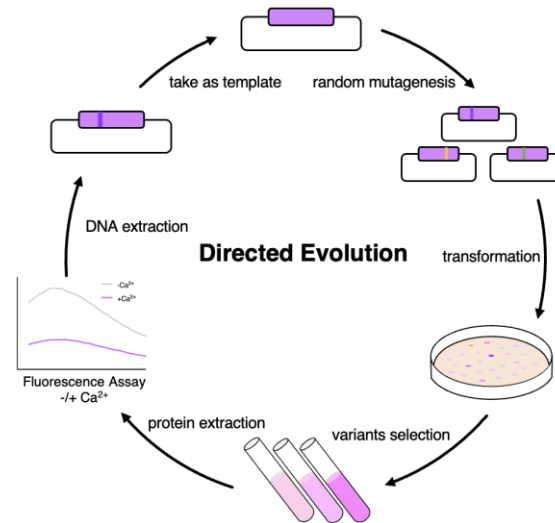
# Development of NIR-GECO3

## 1. Use of brighter FP as a scaffold

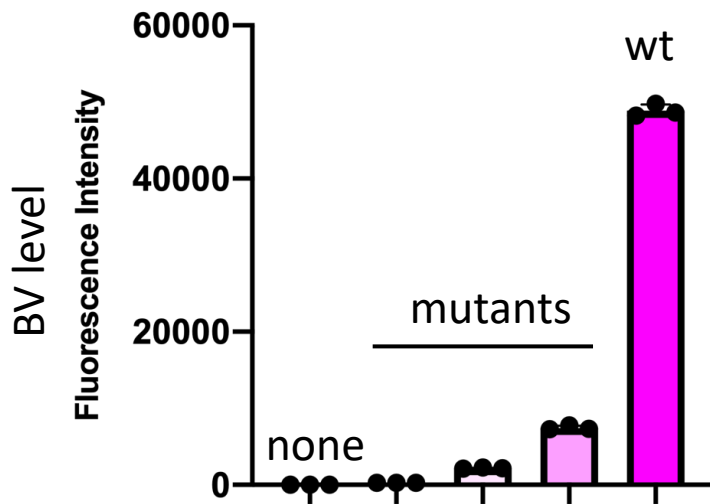


Matlashov et al. *Nat Methods* (2020) 11, 239

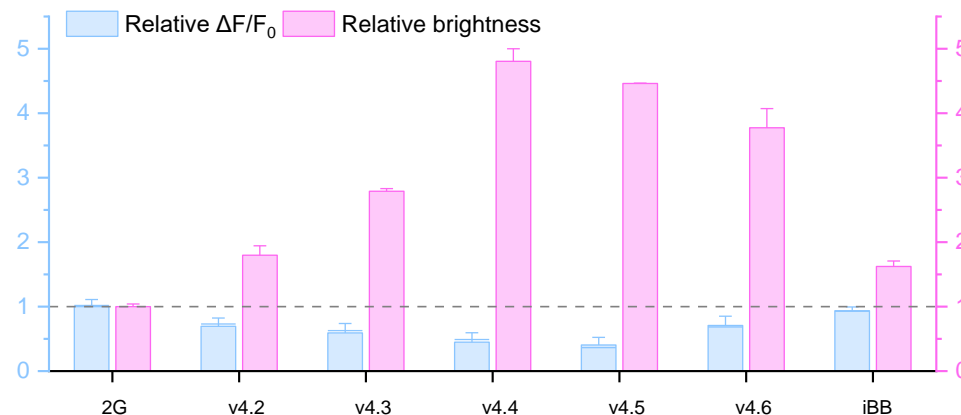
## 2. Directed evolution



## 3. Screening with HO-1 mutant



## 4. Evaluation in mammalian cells

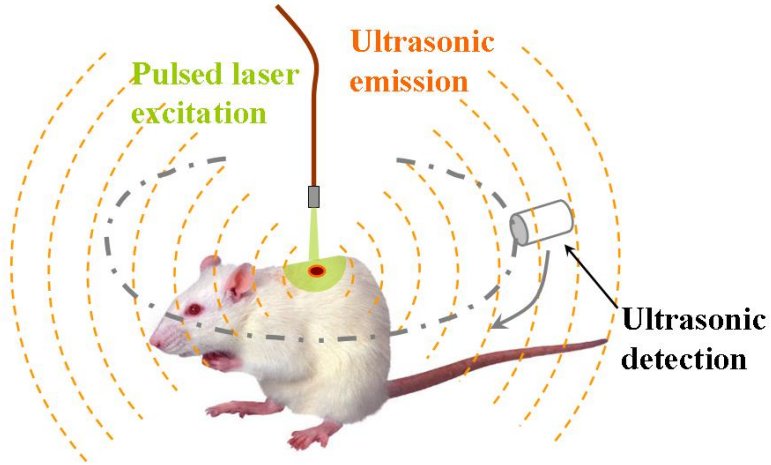


With Prof. Pen Zou  
(Pekin University)



# Optoacoustic imaging of $\text{Ca}^{2+}$

## Optoacoustic imaging



Excitation of light-absorbing molecules



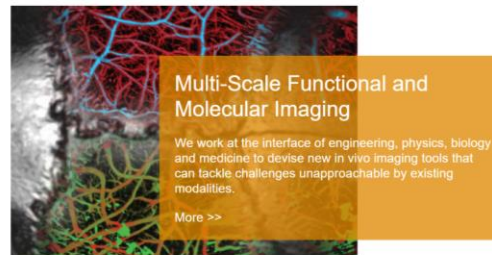
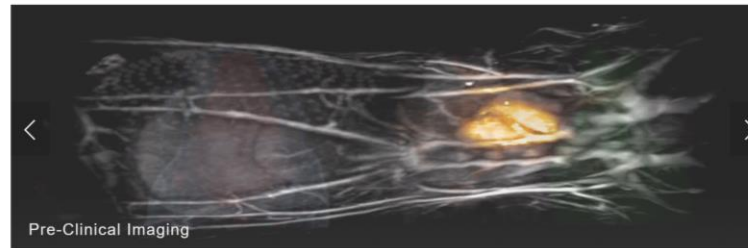
Energy is converted to ultrasound wave

For optoacoustic imaging, the sensor should be non-fluorescent but change its absorbance, in response to the target. We are derivatizing NIR-GECO for such purpose, in collaboration with the Razansky lab.

## Razansky Lab @ETHZ/UZH



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### Main News

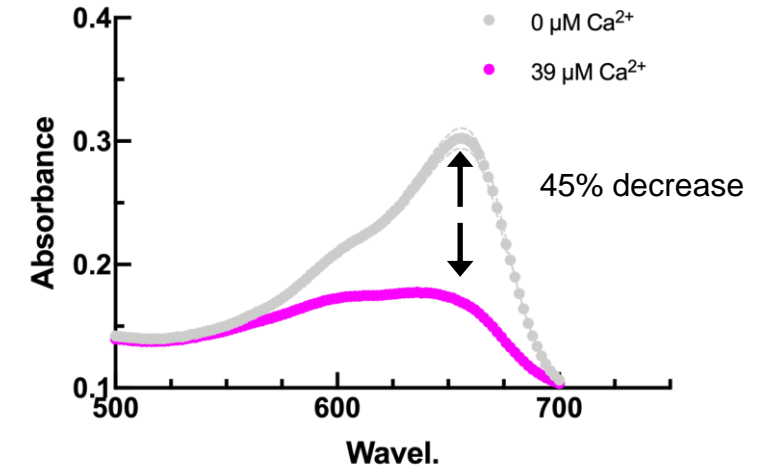
Open webinar at the UCI Beckman Laser Institute »

Optoacoustic-guided focused ultrasound for neurosurgery »

Congratulations to Jaber Malekzadeh for winning the prestigious ETH Fellowship »

We host SONANO focus project together with Hermann lab »

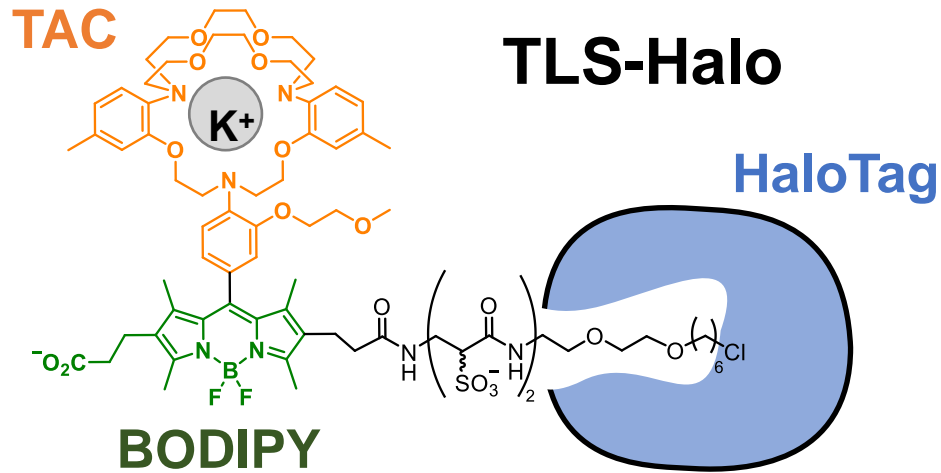
## Absorbance change of NIR-GECO



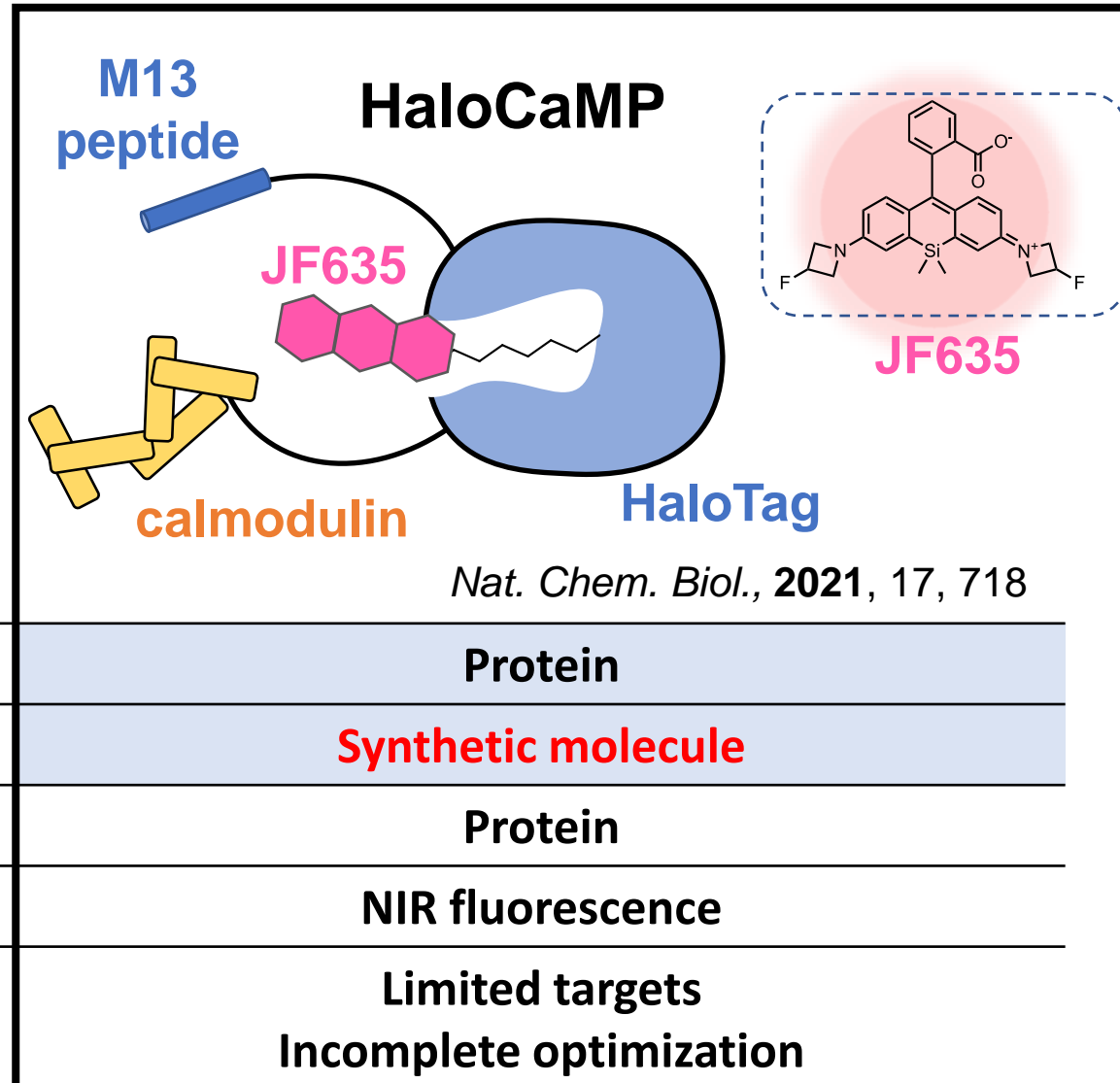
*Change of optoacoustic signal?*

# Chemigenetic sensors

**Chemigenetic** = chemical compound + genetically encoded protein

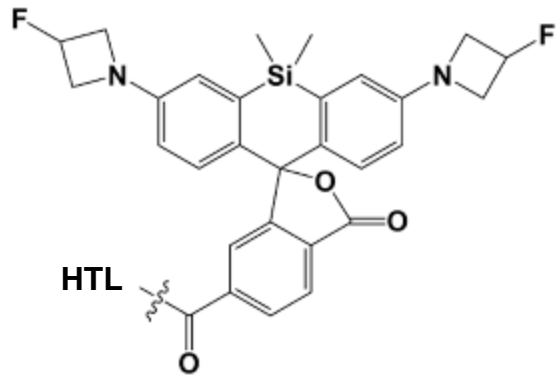


*Anal. Chem.*, **2016**, 88, 2693

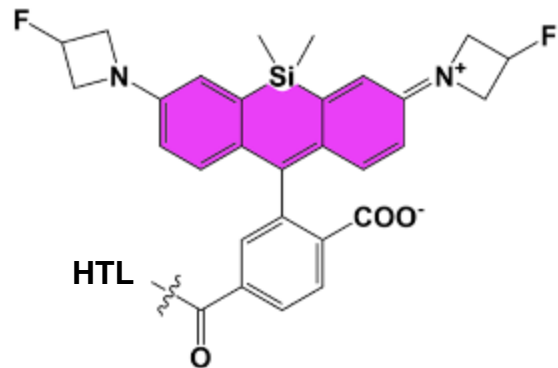


Synthetic molecule	<b>Binder</b>	Protein
Synthetic molecule	<b>Fluorophore</b>	<b>Synthetic molecule</b>
Protein	<b>Scaffold</b>	Protein
Selective localization	<b>Pros</b>	NIR fluorescence
Tedious synthesis No directed evolution	<b>Cons</b>	Limited targets Incomplete optimization

# Chemigenetic K<sup>+</sup> sensor

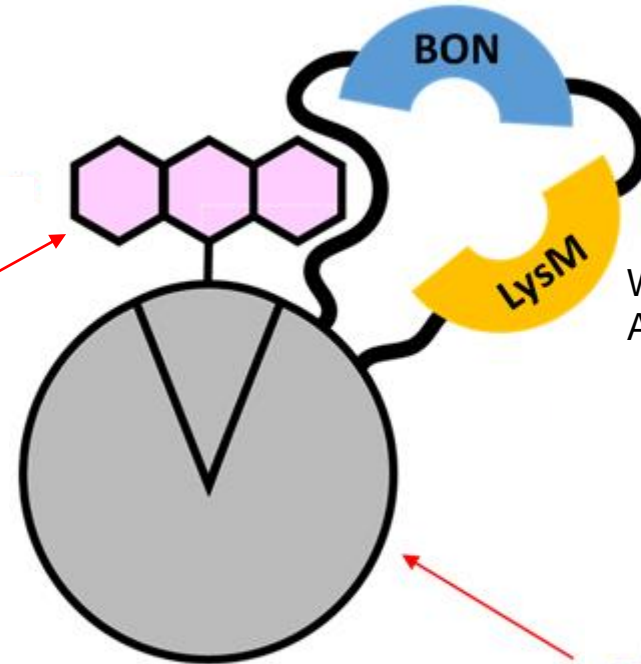


Dim state (Lactone)



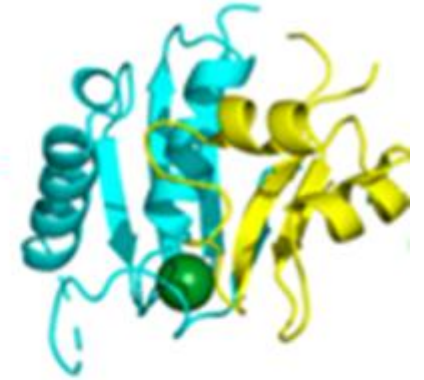
Bright state (Zwitterion)

Janelia Fluor (JF<sub>635</sub>) **Dye** as the fluorophore (+ HTL= HaloTag ligand)

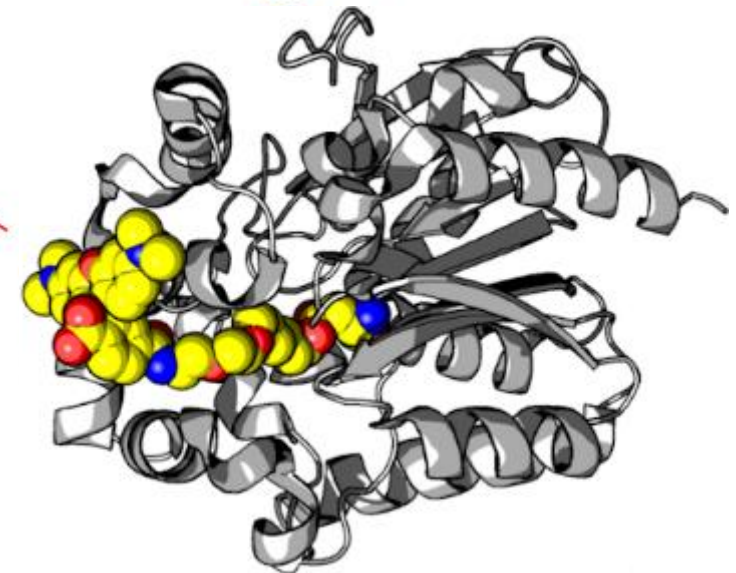


Split HaloTag **Protein** as the scaffold

Kbp **Protein** as the receptor



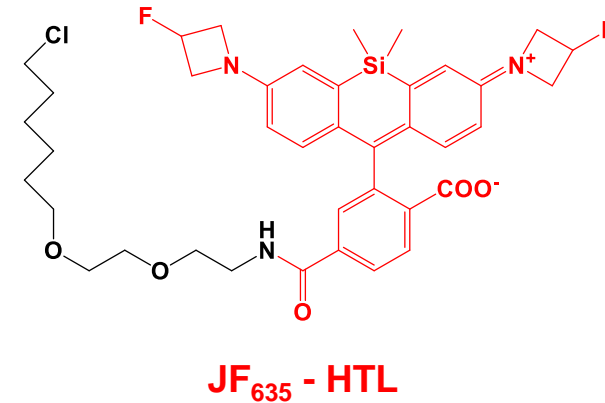
Wu, S. Y., et al, *PLoS Biology*, **2022**, 20, e3001772.  
Ashraf, K. U., et al, *Structure*, **2016**, 24, 741–749.



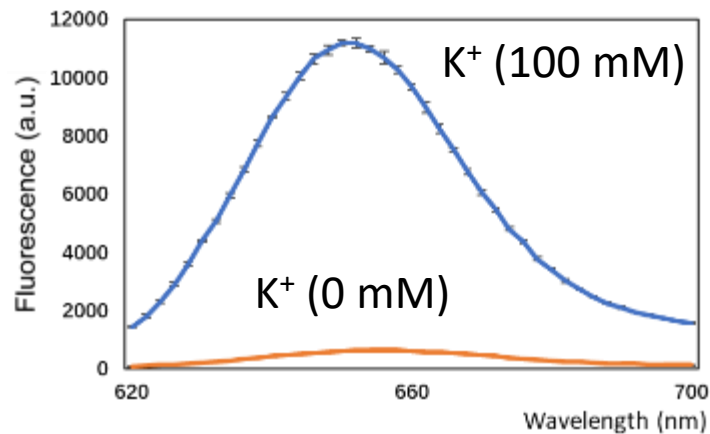


# Performance of the sensor

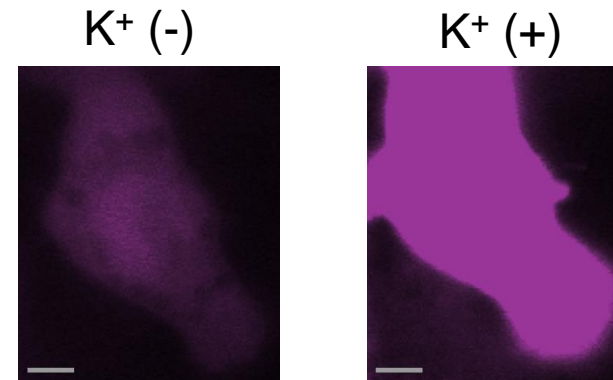
<i>In vitro</i> data (JF <sub>635</sub> )	Halo-Kbp4.2	HaloTag only
$K_d$ (mM)	$35.9 \pm 5.4$	
$\Delta F/F_{0max}$	16.39	-0.02
Quantum Yield [K <sup>+</sup> (100 mM)/K <sup>+</sup> (-)]	0.72/0.52	0.63/0.64



Emission spectra

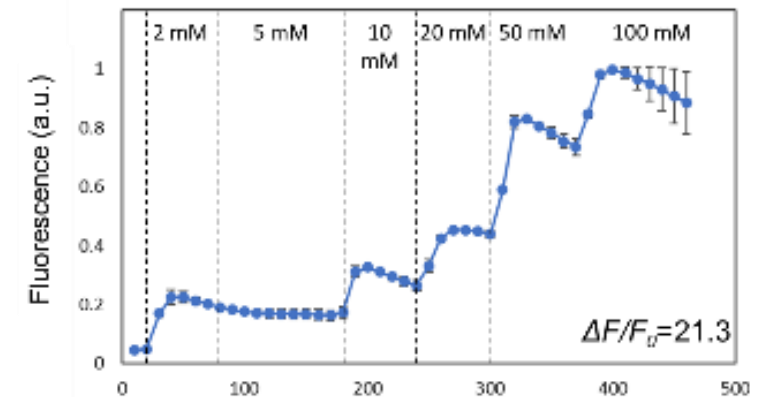


HeLa cell imaging



(digitonin permeabilized)

Timecourse of cell fluorescence



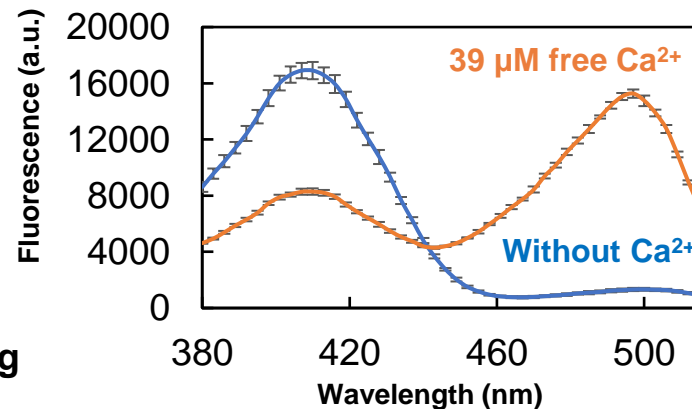
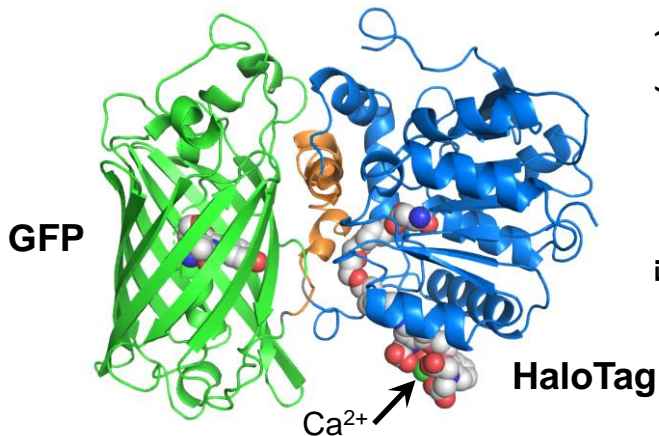
After directed evolution and protein engineering, the sensor showed >15 –fold fluorescence change in response to K<sup>+</sup>, and it worked nicely in the cytosol of cells.

# Summary

- In bio(medical) imaging, we often need molecular sensors (wetware).
- Fluorescent sensors based on proteins are useful tools to visualize targets in living cells or experimental animals.
- Directed evolution is a powerful technique to develop high-performance sensors.
- Chemigenetic approach is promising to create new interesting sensors.

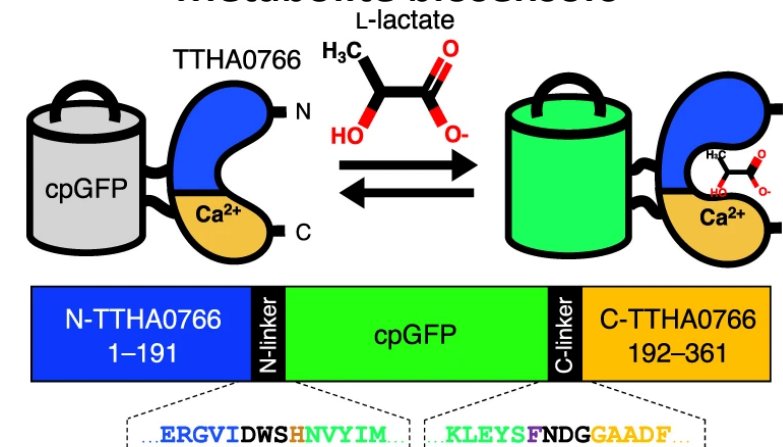
## Other recent work in our lab

### Another type of chemigenetic sensors



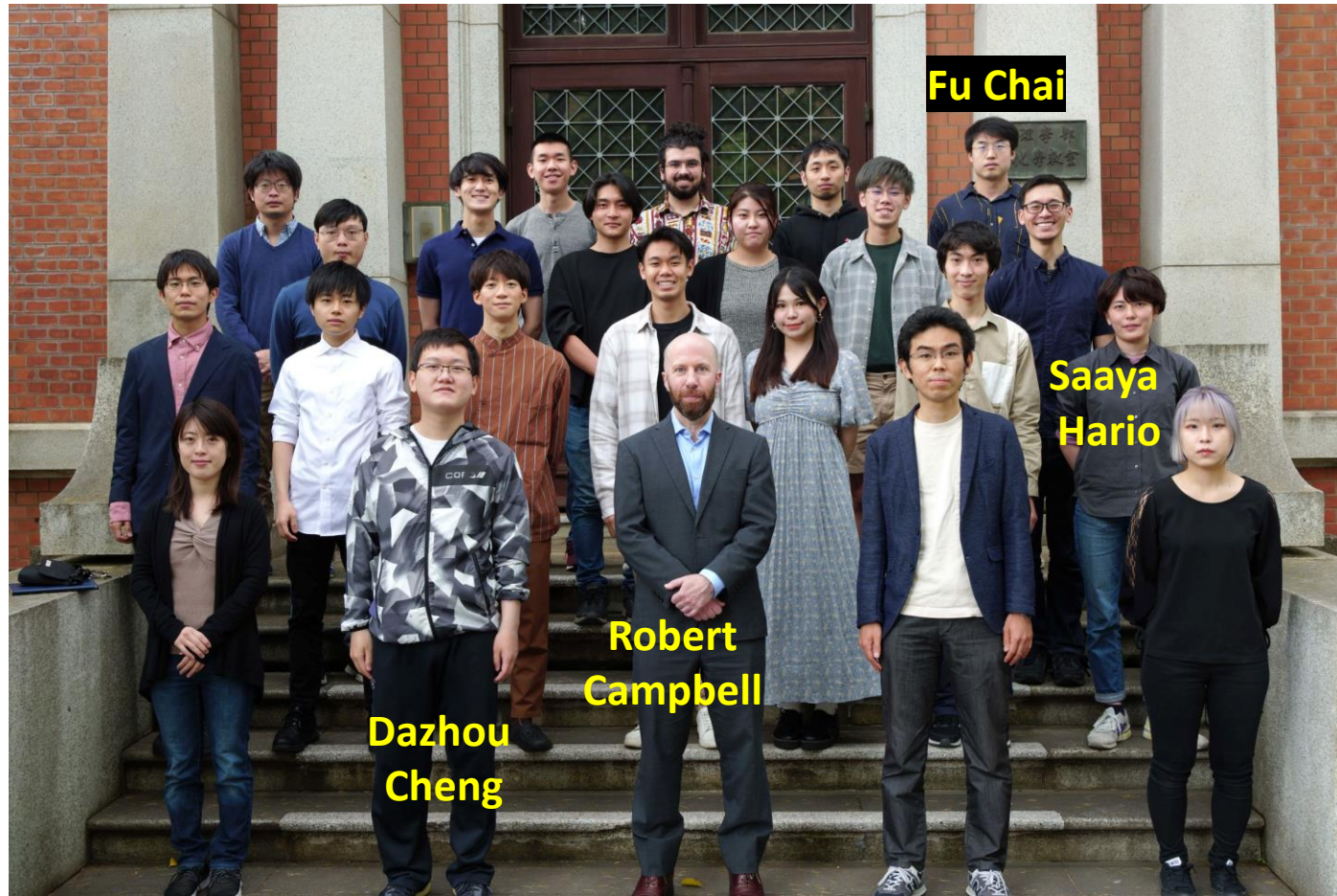
*Nat. Chem. Biol.*, **2023**, 19, 38

### Metabolite biosensors

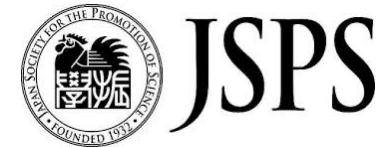


*Nat. Commun.*, **2021**, 12, 7058

# Acknowledgements



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Partnership



Collaborators: Daniel Razansky (ETHZ/UZH), Pen Zou (Peking University), and others